

## Review on Surface Elements and Bacterial Biofilms in Plant-Bacterial Associations

Parwiz Niazi<sup>1</sup>, Abdul Wahid Monib<sup>2</sup>, Hamidullah Ozturk<sup>3</sup>, Mujibullah Mansoor<sup>4</sup>, Azizaqa Azizi<sup>5</sup> and Mohammad Hassan Hassand<sup>6</sup>

<sup>1</sup>Department of Biology, Faculty of Education, Kandahar University, Kandahar, AFGHANISTAN and Department of Plant Protection, Faculty of Agriculture, Ege University, Izmir, TURKEY.

<sup>2</sup>Department of Biology, Faculty of Education, Kandahar University, Kandahar, AFGHANISTAN and School of Environmental Sciences, Jawaharlal Nehru University, New Delhi, INDIA.

<sup>3</sup>Department of Agronomy, Faculty of Agriculture, Faryab University, Faryab, AFGHANISTAN.

<sup>4</sup>Department of Horticulture, Faculty of Agriculture, EGE University, Izmir, TURKEY.

<sup>5</sup>Department of Biology, Faculty of Education, Parwan University, Parwan, AFGHANISTAN.

<sup>6</sup>Department of Biology, Faculty of Education, Kandahar University, Kandahar, AFGHANISTAN.

<sup>1</sup>Corresponding Author: parwiz60@gmail.com



www.jrasb.com || Vol. 2 No. 1 (2023): February Issue

Received: 30-01-2023

Revised: 20-02-2023

Accepted: 02-03-2023

### ABSTRACT

In recent years, there has been increasing interest in the function of bacterial surface elements and functional signals in biofilm formation. Plant-associated bacteria can significantly affect the health and productivity of a plant because they are found in many different areas of the plant, including roots, transport channels, stems, and leaves. The management of these compounds by plants is still unknown, although biofilm production on plants is associated with both symbiotic and pathogenic responses. While some of the bacteria found in biofilm matrices trigger pathogenesis, others can promote plant thriving and serve as biocontrol agents for phytopathogens. This detailed review discusses in depth the various elements and methods involved in the production of bacterial biofilms on plant surfaces and their attachment, as well as the relationship between these factors and bacterial activity and survival.

**Keywords-** Biofilms, Autoaggregation, Plant bacteria, Surface compounds, Exopolymeric compounds.

### I. INTRODUCTION

Bacterial biofilms and surface components play a critical role in the formation and regulation of plant-bacterial associations. These associations are essential for plant growth, health, and productivity (Patwardhan, S. B., et al., 2022). Biofilm formation can promote either symbiotic or pathogenic responses, depending on the bacterial strains involved and the interactions with plants. Therefore, understanding the mechanisms of biofilm formation and attachment to plant surfaces is critical for identifying the key factors that influence

these associations (Beattie, G. A. 2006, Frey-Klett, P., et al., 2007, Trivedi, P., et al., 2020). Biofilm development in plant-bacterial communities has been studied in relation to bacterial surface chemicals, autoaggregation, and exopolymeric substances. For instance, it has been demonstrated that bacterial exopolysaccharides encourage the production of biofilms and their adhesion to plant surfaces (Flemming et al., 2016). There is evidence that bacterial autoaggregation encourages the development of biofilms on plant roots (Giaouris et al., 2015). Also, it has been discovered that the particular surface chemicals produced by bacteria affect their

capacity to colonize plant surfaces and create a biofilm (Lugtenberg and Kamilova, 2009). Biofilm formation on plant surfaces is a complex process involving a series of coordinated steps, including attachment, colonization, and maturation (Liu et al., 2017). The first step of biofilm formation is the reversible attachment of bacteria to the plant surface, which is mediated by bacterial adhesins and plant surface-specific molecules (Ramey et al., 2004). After attachment, bacteria begin to produce and secrete EPS, which helps anchor cells to the surface and provides protection from environmental stresses (O'Toole et al., 2000). Bacterial surface components such as flagella, pili, and lipopolysaccharides are also important for biofilm formation and attachment to plant surfaces (Danhorn and Fuqua, 2007). For example, bacterial flagella play a critical role in initial attachment to plant surfaces as well as in subsequent stages of biofilm development (Guvener and Harwood, 2007). Pili, on the other hand, are involved in the formation of microcolonies and the development of mature biofilms (Beloin et al., 2008). Lipopolysaccharides, the major components of the outer membrane of gram-negative bacteria, have also been shown to play a role in the initial attachment and subsequent maturation of biofilms (Wozniak et al., 2009). Another important factor in biofilm formation is bacterial autoaggregation, which is the spontaneous clumping of bacterial cells (Giaouris et al., 2015). Autoaggregation has been shown to enhance biofilm formation on plant roots by promoting the coaggregation of bacterial cells with each other and with plant surface molecules (Giaouris et al., 2015). In addition, exopolymeric compounds, extracellular polymeric substances produced by bacterial cells, have been shown to promote biofilm formation and attachment to plant surfaces (Flemming et al., 2016). Associations between plants and bacteria can be either beneficial or detrimental to plant health, depending on the bacterial strains involved and the interactions with plants. Some bacteria in biofilms promote plant growth and biocontrol against phytopathogens, while others cause pathogenesis (Lugtenberg and Kamilova, 2009). For example, some bacterial species such as *Pseudomonas fluorescens* and *Bacillus subtilis* have been found to produce compounds that promote plant growth and protect against pathogens. These bacteria are called plant growth-promoting rhizobacteria (PGPR) and are important for maintaining plant health and productivity (Bakker et al., 2013, Niazi, P., 2021). The purpose of this study is to look at how bacterial biofilms and surface elements contribute to plant-bacterial interactions. The proposal concentrates on the processes that lead to the development of biofilms, their attachment to plant surfaces, and the results of these connections on the health and productivity of plants. (Machado, 2020, Nechacov, S. 2019).

## II. BACTERIAL BIOFILMS ON PLANT SURFACES

Bacterial biofilms are complex communities of bacteria that adhere to a surface and produce an extracellular matrix. In the context of plant-microbe interactions, bacterial biofilms can form on various plant surfaces, such as leaves, roots, and seeds. The formation of bacterial biofilms on plant surfaces is a complex process involving the expression of specific genes and the production of various surface components (Avalos, 2020). The first step of bacterial attachment to plant surfaces involves the recognition and interaction of bacterial adhesins with plant cell wall components such as pectin and cellulose (Brewin, N. J. 2004). The expression of motility structures such as flagella and IV-type pili also play a critical role in the early stages of biofilm formation by facilitating bacterial migration to and attachment to the plant surface (Tan, 2016).

Once attached to the surface, bacteria begin to produce an extracellular matrix of polysaccharides, proteins, and lipids that provides structural support and protection for the bacterial community (Friedlander, 2013). The composition and architecture of the matrix can vary depending on the bacterial species and environmental conditions. The biofilm matrix can also serve as a reservoir for nutrients and signaling molecules that facilitate communication and cooperation between bacterial communities (Yaron, 2014).

The formation and structure of bacterial biofilms on plant surfaces have significant implications for plant health and agriculture. For example, bacterial biofilms can protect pathogenic bacteria from plant defense mechanisms and antimicrobial treatments, leading to plant diseases. On the other hand, some beneficial bacteria form biofilms that promote plant growth and nutrient uptake (Muhammad, M. H., 2022, Kolter, R., 2006).

## III. BACTERIAL AUTOAGGREGATION VIA CELL-CELL ADHESION

Bacterial autoaggregation refers to the process by which bacterial cells interact and adhere to each other to form clusters or aggregates. This phenomenon is often mediated by cell surface proteins and other macromolecules that promote adhesive cell-cell interactions (Bogino, P. C., 2013). Autoaggregation can confer several benefits to bacteria, such as increased resistance to environmental stress, enhanced nutrient uptake, and improved virulence. In some cases, autoaggregation may also facilitate the formation of bacterial biofilms, i.e., surface-associated bacterial communities that exhibit increased tolerance to antimicrobial agents and host immune defenses (Nwaiwu, O. 2022).

The molecular mechanisms underlying bacterial autoaggregation are diverse and vary depending on

bacterial species and environmental conditions. Some bacteria utilize surface structures such as pili, fimbriae, and extracellular appendages to promote cell-cell interactions. Other bacteria produce extracellular polysaccharides and other biomolecules that mediate cell-surface interactions (Anburajan, P., 2021). The study of bacterial autoaggregation has significant implications for understanding the basic biology of bacteria and developing new strategies to combat bacterial infections. For example, some researchers are investigating the use of anti-autoaggregation compounds as potential therapeutic agents to disrupt bacterial biofilms and reduce the virulence of pathogenic bacteria (Burger, M. 2000). Overall, bacterial autoaggregation is an important phenomenon that reflects the dynamic and complex interactions between bacterial cells and their environment (Gilbert, P., 2002).

**3.1- Bacterial components involved in plant interactions: adhesins, flagella, pili, and lipopolysaccharides:**

Bacterial surface components play a critical role in establishing and maintaining interactions between plants and bacteria. These components include adhesins, flagella, pili, and lipopolysaccharides (LPS). Adhesins are proteins that allow bacteria to bind to plant surfaces and can be specific to certain types of plant cell wall

components (Pinski, A., 2019). Flagella and pili are appendages that allow bacteria to move on and adhere to plant surfaces. LPS are molecules present in the outer membrane of Gram-negative bacteria that can interact with plant receptors to facilitate bacterial colonization (Pinski, A., 2019). The research looked at the function of particular bacterial surface elements in interactions between plants and bacteria. For instance, it has been demonstrated that the adhesin FimH is essential for the attachment of *Escherichia coli* to plant roots (Lebeis et al., 2015). Plant receptors identify the flagellin protein Flg22, which can start defense processes against bacterial infections (Felix et al., 1999). It has been established that *Xylella fastidiosa's* pili play a role in the bacterium's adhesion to plant xylem vessels and the emergence of Pierce's disease (Chatterjee et al., 2008). It has been demonstrated that *Ralstonia solanacearum* LPS is involved in the colonization of plant roots and the emergence of bacterial wilt disease (Lowe-Power et al., 2016). In general, bacteria's surface elements play a key role as mediators in plant-bacterial interactions, and their effects on agriculture and plant health can be profound (Berlec, A. 2012). Table 1 summarizes some of the bacterial surface components involved in plant-bacterial interactions, their functions, and examples of bacterial species that possess these components.

**Table 1: Bacterial surface components involved in plant-bacterial interactions**

Surface component	Function	Examples of bacterial species
Pili	Promote bacterial attachment to plant surface and facilitate biofilm formation	<i>Escherichia coli</i> , <i>Xylella fastidiosa</i> , <i>Salmonella enterica</i>
Flagella	Facilitate bacterial motility towards and attachments to plant surfaces	<i>Azospirillum brasilense</i> , <i>Rhizobium leguminosarum</i> , <i>Pseudomonas syringae</i>
Adhesins	Mediate bacterial attachments to plant surface	<i>Pseudomonas syringae</i> , <i>Agrobacterium tumefaciens</i> , <i>Xanthomonas campestris</i>
LPS	Interact with plant receptors to facilitate bacterial colonization	<i>Pseudomonas syringae</i> , <i>Xanthomonas campestris</i> , <i>Ralstonia solanacearum</i>

**3.2- Bacterial surface factors in autoaggregation :**

Bacterial autoaggregation is a process in which bacteria bind together through cell-cell interactions. This process can be mediated by various surface factors, including adhesins, pili, fimbriae, extracellular polymeric substances (EPS), and lipopolysaccharides (LPS). Understanding the surface factors involved in bacterial autoaggregation can shed light on the mechanisms underlying biofilm formation and bacterial virulence (Chatterjee, S., 2008, Bogino, P. C., 2013). Several surface variables' effects on bacterial autoaggregation have been the subject of studies. FimH, an adhesin, has been demonstrated to participate in cell-

cell communication and autoaggregation in *Escherichia coli*, for instance (Wellens et al., 2013). Moreover, autoaggregation and the development of biofilms have been linked to the pili of *Enterococcus faecalis* (Shankar et al., 1999). For bacterial autoaggregation and biofilm development in *Pseudomonas aeruginosa*, EPS synthesis has been demonstrated to be essential (Rybtke et al., 2015). LPS has promoted cell-cell contacts and autoaggregation in *Salmonella enterica* (Prouty et al., 2002). Table 2 summarizes some of the surface factors involved in bacterial autoaggregation, along with their functions and examples of bacterial species that possess these factors.

**Table 2: Surface factors involved in bacterial autoaggregation**

Surface factor	Function	Examples of bacterial species
Lipopolysaccharides (LPS)	Interact with other bacteria and promote autoaggregation	<i>Salmonella enterica</i> , <i>Pseudomonas aeruginosa</i> , <i>Escherichia coli</i>

<b>Adhesins</b>	Mediate bacterial attachment to other bacteria	<i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Pseudomonas aeruginosa</i>
<b>Extracellular polymeric substances (EPS)</b>	Facilitate bacterial cell-cell interactions and biofilm formation	<i>Pseudomonas aeruginosa</i> , <i>Staphylococcus epidermidis</i> , <i>Bacillus subtilis</i>
<b>Pili/Fimbriae</b>	Promote bacterial attachment to other bacteria and surfaces	<i>Enterococcus faecalis</i> , <i>Streptococcus mutans</i> , <i>Actinomyces naeslundii</i>

**3.3- Extracellular factors Involved in Bacterial Autoaggregation:**

Bacterial autoaggregation is a process in which bacteria aggregate through cell-cell interactions, which can be mediated by various extracellular factors. Extracellular factors involved in bacterial autoaggregation include extracellular polymeric substances (EPS), quorum-sensing molecules, outer membrane vesicles, and secreted proteins. Understanding the roles of these extracellular factors can provide insight into the mechanisms underlying biofilm formation and bacterial virulence (Vander Hoogerstraete, T., 2013). The research examined the functions of several extracellular elements in bacterial autoaggregation. For instance, it has been demonstrated that EPS synthesis in

*Pseudomonas aeruginosa* is essential for bacterial autoaggregation and biofilm formation (Hultqvist, L. D., et al., 2015). Moreover, it has been demonstrated that quorum sensing molecules, such as N-acyl homoserine lactones (AHLs), are crucial for the autoaggregation and biofilm formation of *Vibrio cholera* and *Pseudomonas aeruginosa* (Ismail, A.S., et al., 2016). It has been demonstrated that secreted proteins from *Staphylococcus aureus* have a role in bacterial autoaggregation and biofilm formation, Examples of these proteins include protein A and biofilm-associated protein (Chen, C., et al., 2001). Table 3 summarizes some of the extracellular factors involved in bacterial autoaggregation, along with their functions and examples of bacterial species that possess these factors.

**Table 3: Extracellular factors involved in bacterial autoaggregation**

<b>Extracellular factor</b>	<b>Function</b>	<b>Examples of bacterial species</b>
<b>Extracellular polymeric substances (EPS)</b>	Facilitate bacterial cell-cell interactions and biofilm formation	<i>Pseudomonas aeruginosa</i> , <i>Staphylococcus epidermidis</i> , <i>Bacillus subtilis</i>
<b>Quorum-sensing molecules</b>	Regulate bacterial gene expression and promote cell-cell communication	<i>Vibrio cholerae</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i>
<b>Outer membrane vesicles</b>	Facilitate bacterial communication and virulence	<i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Vibrio cholera</i>
<b>Secreted proteins</b>	Facilitate cell-cell interactions and biofilm formation	<i>Streptococcus mutans</i> , <i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i>

**IV. BACTERIAL BIOFILM FORMATION VIA CELL-CELL AND CELL-SURFACE INTERACTIONS**

Bacterial biofilm formation is a complex process that involves various cell-cell and cell-surface interactions. These interactions are mediated by different bacterial surface components, including adhesins, pili, flagella, and extracellular polymeric substances (EPS). Understanding the roles of these components can provide insights into the mechanisms underlying bacterial biofilm formation and can help in developing strategies to prevent or treat biofilm-related infections (Hori, K., 2010, Van Houdt, R., et al., 2005). The functions of distinct bacterial surface elements in biofilm formation have been the subject of numerous investigations. The Type IV pili, for instance, are crucial for bacterial adhesion to surfaces and cell-cell interactions during biofilm development in

*Pseudomonas aeruginosa* (Eid, J., et al., 2009). The extracellular matrix, composed of EPS, has also been shown to be important for biofilm formation in *Staphylococcus epidermidis* and *Pseudomonas aeruginosa* (Kaplan, 2010). Adhesins, such as FimH, have been shown to be involved in bacterial attachment and biofilm formation in uropathogenic *Escherichia coli* (Wurpel et al., 2014). Flagella have been shown to be important for motility and biofilm formation in *Vibrio cholerae* (Watnick, et al., 1999). Overall, cell-cell and cell-surface interactions play critical roles in bacterial biofilm formation, and understanding the mechanisms underlying these interactions can lead to the development of new strategies for the prevention and treatment of biofilm-related infections (Habimana, O., et al., 2014). Table 4 summarizes some of the key cell-cell and cell-surface interactions involved in bacterial biofilm formation, along with their functions and examples of bacterial species that possess these components.

**Table 4: Cell-cell and cell-surface interactions involved in bacterial biofilm formation**

Surface component	Function	Examples of bacterial species
<b>Adhesins</b>	Facilitate bacterial attachment to surfaces	<i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Streptococcus mutans</i>
<b>Pili</b>	Facilitate bacterial attachment to surfaces and cell-cell interactions	<i>Pseudomonas aeruginosa</i> , <i>Neisseria gonorrhoeae</i> , <i>Escherichia coli</i>
<b>Flagella</b>	Facilitate bacterial motility and biofilm formation	<i>Pseudomonas aeruginosa</i> , <i>Vibrio cholerae</i> , <i>Escherichia coli</i>
<b>Extracellular polymeric substances (EPS)</b>	Facilitate cell-cell and cell-surface interactions and biofilm formation	<i>Staphylococcus epidermidis</i> , <i>Pseudomonas aeruginosa</i> , <i>Bacillus subtilis</i>

**4.1- Structural and Functional Components Involved in Biofilm Formation:**

Biofilms are complex communities of microorganisms that are attached to surfaces and embedded in a self-produced extracellular matrix. Biofilm formation is a highly regulated process that involves the coordination of various structural and functional components (Vasudevan, R. 2014). The functions of several structural and functional elements in the creation of biofilms have been the subject of numerous research, for example, in *Staphylococcus aureus*, the cell wall-associated adhesins are critical for attachment to surfaces and biofilm formation (Foster, 2019). In *Pseudomonas aeruginosa*, the Type IV pili are essential for bacterial attachment to surfaces and cell-cell interactions during biofilm formation (Ma et al., 2009). The extracellular matrix, composed of EPS, has also been shown to be important for biofilm formation in *Staphylococcus epidermidis* and *Pseudomonas aeruginosa* (Kaplan, 2010, Flemming et al, 2010).

Quorum sensing systems, which regulate gene expression and coordinate biofilm formation, have been shown to be important in *Vibrio cholera* and *Pseudomonas aeruginosa* (Waters and Bassler, 2005; Schuster and Müh, U., 2006). Enzymes, such as proteases and DNases, have been shown to contribute to biofilm dispersal in *Staphylococcus aureus* and *Pseudomonas aeruginosa* (Boles and Horswill, 2011; Flemming and Wingender, 2010). Overall, the structural and functional components involved in biofilm formation are diverse and complex, and their roles are still being elucidated. However, understanding the mechanisms underlying biofilm formation and the roles of specific components can lead to the development of new strategies for the prevention and treatment of biofilm-related infections (Madsen, J. S., et al 2012). Table 5 summarizes some of the key structural and functional components involved in biofilm formation, along with their functions and examples of bacterial species that possess these components.

**Table 5: Structural and functional components involved in biofilm formation**

Component	Function	Examples of bacterial species
<b>Adhesins</b>	Mediate attachment to surfaces	<i>Staphylococcus aureus</i> , <i>Pseudomonas aeruginosa</i> , <i>Escherichia coli</i>
<b>Pili</b>	Mediate attachment to surfaces and facilitate cell-cell interactions	<i>Neisseria gonorrhoeae</i> , <i>Pseudomonas aeruginosa</i> , <i>Escherichia coli</i>
<b>Extracellular polymeric substances (EPS)</b>	Mediate cell-cell and cell-surface interactions and provide structural support	<i>Staphylococcus epidermidis</i> , <i>Pseudomonas aeruginosa</i> , <i>Bacillus subtilis</i>
<b>Quorum sensing systems</b>	Regulate gene expression and coordinate biofilm formation	<i>Vibrio cholerae</i> , <i>Pseudomonas aeruginosa</i> , <i>Streptococcus pneumoniae</i>
<b>Enzymes</b>	Degrade host tissue and contribute to biofilm dispersal	<i>Staphylococcus aureus</i> , <i>Streptococcus mutans</i> , <i>Pseudomonas aeruginosa</i>

**4.2- Extracellular Factors:**

Extracellular factors are molecules that are secreted by bacterial cells and play important roles in various biological processes such as biofilm formation and host-pathogen interactions (Herkert, P. F., et al., 2019). Table 6 summarizes some of the important extracellular factors and their functions. Extracellular polysaccharides (EPS) are important for providing structural integrity and protection against environmental stresses (D'aes, J., et al., 2010). Alginate is an EPS

produced by *Pseudomonas aeruginosa* that contributes to biofilm formation and protects the bacteria from host immune responses (Jeong, J., et al., 2014). Extracellular DNA (eDNA) is another factor that can promote biofilm formation and mediate horizontal gene transfer. DNABII proteins are eDNA-binding proteins that contribute to the structural integrity of bacterial biofilms (Goodman et al., 2011).

Quorum-sensing molecules are small signaling molecules that mediate cell-to-cell communication and

regulate gene expression. Acyl-homoserine lactones (AHLs) are quorum-sensing molecules produced by many Gram-negative bacteria that are involved in various biological processes such as biofilm formation and virulence (Hartmann, A., et al., 2012). Enzymes are another group of extracellular factors that are important for bacterial pathogenesis. Proteases and lipases can degrade host tissues and facilitate nutrient acquisition. For example, the protease SpeB produced by *Streptococcus pyogenes* can degrade host immune molecules and facilitate bacterial infection (Morgan, P. J., et al., 2013).

Toxins are another important group of extracellular factors that can damage host cells and

evade host immune responses. Pore-forming toxins, such as  $\alpha$ -toxin produced by *Staphylococcus aureus*, can damage host cells by creating pores in the cell membrane (Wilke and Bubeck Wardenburg, 2010). Exotoxins are proteins that are secreted by bacteria and can cause various types of damage to host cells. For example, the exotoxin A produced by *Pseudomonas aeruginosa* can inhibit protein synthesis in host cells and contribute to bacterial virulence (Kabir et al., 2016). Overall, extracellular factors play important roles in bacterial pathogenesis and interaction with the host. Understanding these factors can lead to the development of new strategies for the prevention and treatment of bacterial infections (Wilson, J. W., et al., 2002).

**Table 6: Extracellular factors and their functions**

Factor	Function	Examples
Extracellular polysaccharides (EPS)	Provide structural integrity and protect against environmental stresses	<i>Alginate in Pseudomonas aeruginosa</i>
Extracellular DNA (eDNA)	Promote biofilm formation and mediate horizontal gene transfer	DNABII proteins
Quorum-sensing molecules	Regulate gene expression and mediate cell-to-cell communication	<i>Acyl-homoserine lactones (AHLs)</i>
Enzymes	Degrade host tissues and facilitate nutrient acquisition	<i>Proteases, lipases</i>
Toxins	Damage host cells and evade host immune responses	<i>Pore-forming toxins, exotoxins</i>

## V. RELATIONSHIP BETWEEN BIOFILM FORMATION AND BACTERIAL AUTOAGGREGATION

Biofilm formation and bacterial autoaggregation are two closely related processes that play crucial roles in bacterial adhesion and colonization on various surfaces, including biotic and abiotic surfaces (Trunk, T., et al., 2018). Autoaggregation is the process of bacterial self-aggregation, which is mediated by specific cell-surface adhesion factors such as pili, fimbriae, and extracellular polymeric substances (EPS). On the other hand, biofilm formation is a complex process that involves the attachment of bacteria to a

surface, followed by the production of EPS and the formation of a three-dimensional structure (Sorroche, F. G., et al., 2012).

There is a close relationship between bacterial autoaggregation and biofilm formation. Autoaggregation plays a crucial role in the initial stages of biofilm formation by promoting bacterial adhesion and facilitating the formation of microcolonies (Karygianni, L., et al., 2020). Moreover, autoaggregation has been found to be essential for the formation of mature biofilms, as it allows the bacteria to remain in close proximity, promoting cell-to-cell signaling, and EPS production (Wang, H., et al., 2013).

**Table 7: Examples of bacterial surface factors involved in autoaggregation and biofilm formation**

Surface factor	Function in autoaggregation	Function in biofilm formation
Pili	Mediate cell-to-cell adhesion	Promote initial bacterial attachment
Extracellular polymeric substances (EPS)	Facilitate bacterial aggregation	Contribute to the formation of the extracellular matrix and three-dimensional structure
Curli fibers	Promote bacterial self-aggregation	Promote initial bacterial attachment and stability of mature biofilms
Fimbriae	Mediate cell-to-cell adhesion	Promote initial bacterial attachment and facilitate the formation of micro colonies
Lipopolysaccharides (LPS)	Mediate bacterial adhesion and autoaggregation	Promote initial bacterial attachment and contribute to the formation of the extracellular matrix

## VI. ROLE OF BACTERIAL BIOFILMS AND SURFACE COMPONENTS IN PLANT-BACTERIAL ASSOCIATIONS: COLONIZATION AND INFECTION

Bacterial biofilms and surface components play a critical role in the formation and maintenance of plant-bacteria associations. These interactions can be mutualistic, commensal, or pathogenic and are essential for plant growth and development, nutrient cycling, and disease resistance (Venturi, V., & Bez, C. 2021). Interactions between plants and bacteria begin with bacterial adhesion to the plant surface, which is mediated by various bacterial surface components such as

adhesins, flagella, pili, and lipopolysaccharides. Once attached, bacteria can form biofilms on the plant surface that protect them from environmental stresses and promote bacterial survival and growth. In addition, biofilms provide a physical barrier that prevents colonization by competing bacterial species (Wheatley, R. M., 2018, Ahmad, I., et al., 2017).

The establishment and maintenance of plant-bacterial associations also involve communication between the bacteria and the plant, which is mediated by a variety of signaling molecules, such as quorum-sensing molecules, phytohormones, and volatile organic compounds. These signaling molecules play a critical role in regulating plant growth and development, as well as modulating plant immune responses (Beattie, G. A. 2006).

**Table 8: Examples of bacterial surface components involved in plant-bacterial interactions**

Surface component	Function in plant-bacterial interactions
<b>Adhesins</b>	Mediate bacterial adhesion to plant surfaces
<b>Flagella</b>	Facilitate bacterial motility and penetration of plant tissues
<b>Pili</b>	Mediate cell-to-cell adhesion and promote bacterial aggregation
<b>Lipopolysaccharides (LPS)</b>	Mediate bacterial adhesion to plant surfaces and modulate plant immune responses
<b>Extracellular polymeric substances (EPS)</b>	Contribute to the formation of biofilms and protect bacteria from environmental stresses

## VII. IMPACT OF BACTERIAL BIOFILMS ON PLANTS: GROWTH, DISEASE RESISTANCE, AND STRESS TOLERANCE

Plant-associated bacterial biofilms have a significant impact on plant growth, development, disease resistance, and stress tolerance. These biofilms can

influence various physiological and biochemical processes in the plant and facilitate the uptake of nutrients and water (Hashem, A., et al., 2019, Kasim, W. A., et al., 2016). The following table provides some examples of how plant-associated bacterial biofilms impact plant growth and development, disease resistance, and stress tolerance.

**Table 9: Impact of plant-associated bacterial biofilms on plant growth and development, disease resistance, and stress tolerance**

Effect of bacterial biofilms on plant	Example of mechanism
<b>Promotion of plant growth and development</b>	Improving nutrient availability through fixation, solubilization and chelation of nutrients Production of phytohormones that stimulate plant growth and development
<b>Protection against plant pathogens and diseases</b>	Production of antibiotics that inhibit growth of plant pathogens Induction of systemic resistance in plants through the activation of defense mechanisms
<b>Enhancement of plant stress tolerance</b>	Production of compounds that enhance plant tolerance to drought, salinity, and heavy metal stress Improving photosynthetic efficiency under stress conditions

### VIII. MANIPULATING BACTERIAL BIOFILMS AND COMPONENTS TO IMPROVE PLANT-MICROBE INTERACTIONS IN AGRICULTURE AND ENVIRONMENT

Manipulating bacterial biofilms and surface constituents can be an effective strategy to improve

plant-microbe interactions for various agricultural and environmental applications. This can include the use of beneficial bacteria to promote plant growth and development, the development of biofilms to protect against plant pathogens and environmental stresses, and the use of biofilms to bio-remediate contaminated soils and waters (Adak, A., et al. 2016, Berg, G. 2009). The following table provides some examples of strategies for manipulating bacterial biofilms and surface components to enhance plant-microbe interactions.

**Table 10: Strategies for manipulating bacterial biofilms and surface components for agricultural and environmental applications**

Manipulation strategy	Example
<i>Introduction of beneficial bacteria</i>	Inoculating plant roots with nitrogen-fixing bacteria to enhance plant growth and reduce reliance on chemical fertilizers. Using plant growth-promoting bacteria to enhance nutrient availability and promote stress tolerance
<b>Development of protective biofilms</b>	Engineering bacteria to produce antimicrobial compounds that inhibit plant pathogens. Manipulating bacterial adhesins to enhance attachment to plant surfaces and promote biofilm formation.
<b>Use of biofilms for bioremediation</b>	Developing biofilms to degrade pollutants in contaminated soils and water. Using biofilms for the immobilization of heavy metals and other toxic compounds

### IX. CONCLUDING REMARKS

The bacterial cell surface plays a critical role in promoting bacterial spread, survival, and attachment to plant surfaces through autoaggregation and biofilm formation. Both bacterial surface factors and extracellular factors play critical roles in these processes, which are essential for plant colonization. Bacteria living in biofilms benefit from significant advantages such as protection from predators, desiccation, and antibacterial agents. Biofilms serve as a survival habitat for both beneficial and pathogenic bacteria, increasing their potential to survive and thrive in the plant environment. In addition, biofilms have been shown to improve individual bacterial fitness and overall plant health and productivity through cumulative selective benefits. Understanding the molecular mechanisms underlying biofilm formation and microbial adaptations to this life form in a variety of environments is critical to understanding bacterial interactions with their eukaryotic hosts, including plants. The complex interactions between prokaryotes and eukaryotes will be clarified by multidisciplinary studies employing novel methodologies that will assist understand the mechanisms at various phases of biofilm formation on plant surfaces. In the end, this information will open the door for the creation of methods to control bacterial biofilms for use in agricultural and environmental protection.

### REFERENCES

- [1] Ahmad, I., Khan, M. S., Altaf, M. M., Qais, F. A., Ansari, F. A., & Rumbaugh, K. P. (2017). Biofilms: an overview of their significance in plant and soil health. *Biofilms in plant and soil health*, 1-25.
- [2] Adak, A., Prasanna, R., Babu, S., Bidyarani, N., Verma, S., Pal, M., & Nain, L. (2016). Micronutrient enrichment mediated by plant-microbe interactions and rice cultivation practices. *Journal of Plant Nutrition*, 39(9), 1216-1232.
- [3] Anburajan, P., Kim, Y., Rice, S. A., & Oh, H. S. (2021). Bacterial signaling and signal responses as key factors in water and wastewater treatment. *Journal of Water Process Engineering*, 44, 102434.
- [4] Avalos, A. P., Fernández, R. L., & Pérez, D. Z. (2020). Validación de la escala de incapacidad por dolor lumbar de Oswestry, en paciente con dolor crónico de la espalda. Cienfuegos, 2017-2018. *Rehabilitación*, 54(1), 25-30.
- [5] Bakker, A. B., & Sanz-Vergel, A. I. (2013). Weekly work engagement and flourishing: The role of hindrance and challenge job demands. *Journal of Vocational Behavior*, 83(3), 397-409.
- [6] Beattie, G. A. (2006). Plant-associated bacteria: survey, molecular phylogeny, genomics and recent advances. *Plant-associated bacteria*, 1-56.
- [7] Berg, G. (2009). Plant-microbe interactions promoting plant growth and health: perspectives for



controlled use of microorganisms in agriculture. *Applied microbiology and biotechnology*, 84, 11-18.

[8] Berlec, A. (2012). Novel techniques and findings in the study of plant microbiota: search for plant probiotics. *Plant science*, 193, 96-102.

[9] Bogino, P. C., de las Mercedes Oliva, M., Sorroche, F. G., & Giordano, W. (2013). The role of bacterial biofilms and surface components in plant-bacterial associations. *International journal of molecular sciences*, 14(8), 15838-15859.

[10] Brewin, N. J. (2004). Plant cell wall remodelling in the Rhizobium-legume symbiosis. *Critical Reviews in Plant Sciences*, 23(4), 293-316.

[11] Chen, C., Pore, N., Behrooz, A., Ismail-Beigi, F., & Maity, A. (2001). Regulation of glut1 mRNA by hypoxia-inducible factor-1: interaction between H-ras and hypoxia. *Journal of Biological Chemistry*, 276(12), 9519-9525.

[12] Chatterjee, S., Ford, E. B., Matsumura, S., & Rasio, F. A. (2008). Dynamical outcomes of planet-planet scattering. *The Astrophysical Journal*, 686(1), 580.

[13] Burger, M., Woods, R. G., McCarthy, C., & Beacham, I. R. (2000). Temperature regulation of protease in *Pseudomonas fluorescens* LS107d2 by an ECF sigma factor and a transmembrane activator. *Microbiology*, 146(12), 3149-3155.

[14] D'aes, J., De Maeyer, K., Pauwelyn, E., & Höfte, M. (2010). Biosurfactants in plant-Pseudomonas interactions and their importance to biocontrol. *Environmental microbiology reports*, 2(3), 359-372.

[15] Danhorn, T., & Fuqua, C. (2007). Biofilm formation by plant-associated bacteria. *Annu. Rev. Microbiol.*, 61, 401-422.

[16] Friedlander, R. S., Vlamakis, H., Kim, P., Khan, M., Kolter, R., & Aizenberg, J. (2013). Bacterial flagella explore microscale hummocks and hollows to increase adhesion. *Proceedings of the National Academy of Sciences*, 110(14), 5624-5629.

[17] Eid, J., Fehr, A., Gray, J., Luong, K., Lyle, J., Otto, G., & Turner, S. (2009). Real-time DNA sequencing from single polymerase molecules. *Science*, 323(5910), 133-138.

[18] Flemming, H. C., Wingender, J., Szewzyk, U., Steinberg, P., Rice, S. A., & Kjelleberg, S. (2016). Biofilms: an emergent form of bacterial life. *Nature Reviews Microbiology*, 14(9), 563-575.

[19] Flemming, H. C., & Wingender, J. (2010). The biofilm matrix. *Nature reviews microbiology*, 8(9), 623-633.

[20] Felix, G., Duran, J. D., Volko, S., & Boller, T. (1999). Plants have a sensitive perception system for the most conserved domain of bacterial flagellin. *The Plant Journal*, 18(3), 265-276.

[21] Frey-Klett, P., Garbaye, J., & Tarkka, M. (2007). The mycorrhiza helper bacteria revisited. *New phytologist*, 176(1), 22-36.

[22] Foster, K. R., & Schwan, H. P. (2019). Dielectric properties of tissues. *CRC handbook of biological effects of electromagnetic fields*, 27-96.

[23] Giaouris, E., Heir, E., Desvaux, M., Hébraud, M., Møretre, T., Langsrud, S., & Simões, M. (2015). Intra- and inter-species interactions within biofilms of important foodborne bacterial pathogens. *Frontiers in microbiology*, 6, 841.

[24] Gilbert, P., Maira-Litran, T., McBain, A. J., Rickard, A. H., & Whyte, F. W. (2002). The physiology and collective recalcitrance of microbial biofilm communities.

[25] Goodman, S. H., Rouse, M. H., Connell, A. M., Broth, M. R., Hall, C. M., & Heyward, D. (2011). Maternal depression and child psychopathology: A meta-analytic review. *Clinical child and family psychology review*, 14, 1-27.

[26] Güvener, Z. T., & Harwood, C. S. (2007). Subcellular location characteristics of the *Pseudomonas aeruginosa* GGDEF protein, WspR, indicate that it produces cyclic-di-GMP in response to growth on surfaces. *Molecular microbiology*, 66(6), 1459-1473.

[27] Habimana, O., Semião, A. J. C., & Casey, E. (2014). The role of cell-surface interactions in bacterial initial adhesion and consequent biofilm formation on nanofiltration/reverse osmosis membranes. *Journal of Membrane Science*, 454, 82-96.

[28] Hartmann, A., & Schikora, A. (2012). Quorum sensing of bacteria and trans-kingdom interactions of N-acyl homoserine lactones with eukaryotes. *Journal of chemical ecology*, 38, 704-713.

[29] Hashem, A., Tabassum, B., & Abd\_Allah, E. F. (2019). *Bacillus subtilis*: A plant-growth promoting rhizobacterium that also impacts biotic stress. *Saudi journal of biological sciences*, 26(6), 1291-1297.

[30] Herkert, P. F., Amatuzzi, R. F., Alves, L. R., & Rodrigues, M. L. (2019). Extracellular vesicles as vehicles for the delivery of biologically active fungal molecules. *Current Protein and Peptide Science*, 20(10), 1027-1036.

[31] Hori, K., & Matsumoto, S. (2010). Bacterial adhesion: From mechanism to control. *Biochemical Engineering Journal*, 48(3), 424-434.

[32] Hultqvist, L. D., Givskov, M., & Tolker-Nielsen, T. (2015). *Pseudomonas aeruginosa* biofilm infections: community structure, antimicrobial tolerance and immune response. *Journal of molecular biology*, 427(23), 3628-3645.

[33] Ismail, A. S., Valastyan, J. S., & Bassler, B. L. (2016). A host-produced autoinducer-2 mimic activates bacterial quorum sensing. *Cell Host & Microbe*, 19(4), 470-480.

[34] Jeong, J., Kang, H. M., Lee, E. K., Song, B. M., Kwon, Y. K., Kim, H. R., ... & Lee, Y. J. (2014). Highly pathogenic avian influenza virus (H5N8) in domestic poultry and its relationship with migratory birds in South Korea during 2014. *Veterinary microbiology*, 173(3-4), 249-257.

- [35] Kamilova, F., Kravchenko, L. V., Shaposhnikov, A. I., Azarova, T., Makarova, N., & Lugtenberg, B. (2006). Organic acids, sugars, and L-tryptophane in exudates of vegetables growing on stonewool and their effects on activities of rhizosphere bacteria. *Molecular Plant-Microbe Interactions*, 19(3), 250-256.
- [36] Kaplan, J. Á. (2010). Biofilm dispersal: mechanisms, clinical implications, and potential therapeutic uses. *Journal of dental research*, 89(3), 205-218.
- [37] Karygianni, L., Ren, Z., Koo, H., & Thurnheer, T. (2020). Biofilm matrixome: extracellular components in structured microbial communities. *Trends in Microbiology*, 28(8), 668-681.
- [38] Kasim, W. A., Gaafar, R. M., Abou-Ali, R. M., Omar, M. N., & Hewait, H. M. (2016). Effect of biofilm forming plant growth promoting rhizobacteria on salinity tolerance in barley. *Annals of Agricultural Sciences*, 61(2), 217-227.
- [39] Kim, K. H., Kabir, E., & Jahan, S. A. (2016). A review on the distribution of Hg in the environment and its human health impacts. *Journal of hazardous materials*, 306, 376-385.
- [40] Kolter, R., & Greenberg, E. P. (2006). The superficial life of microbes. *Nature*, 441(7091), 300-302.
- [41] Lebeis, Sarah L., Sur Herrera Paredes, Derek S. Lundberg, Natalie Breakfield, Jase Gehring, Meredith McDonald, Stephanie Malfatti et al. "Salicylic acid modulates colonization of the root microbiome by specific bacterial taxa." *Science* 349, no. 6250 (2015): 860-864.
- [42] Lugtenberg, B., & Kamilova, F. (2009). Plant-growth-promoting rhizobacteria. *Annual review of microbiology*, 63, 541-556.
- [43] Machado, I., Silva, L. R., Giaouris, E. D., Melo, L. F., & Simões, M. (2020). Quorum sensing in food spoilage and natural-based strategies for its inhibition. *Food Research International*, 127, 108754.
- [44] Madsen, J. S., Burmølle, M., Hansen, L. H., & Sørensen, S. J. (2012). The interconnection between biofilm formation and horizontal gene transfer. *FEMS Immunology & Medical Microbiology*, 65(2), 183-195.
- [45] Mason, C. J., Lowe-Power, T. M., Rubert-Nason, K. F., Lindroth, R. L., & Raffa, K. F. (2016). Interactions between bacteria and aspen defense chemicals at the phyllosphere-herbivore interface. *Journal of chemical ecology*, 42, 193-201.
- [46] Morgan, P. J., Barnett, L. M., Cliff, D. P., Okely, A. D., Scott, H. A., Cohen, K. E., & Lubans, D. R. (2013). Fundamental movement skill interventions in youth: A systematic review and meta-analysis. *Pediatrics*, 132(5), e1361-e1383.
- [47] Müh, U., Schuster, M., Heim, R., Singh, A., Olson, E. R., & Greenberg, E. P. (2006). Novel *Pseudomonas aeruginosa* quorum-sensing inhibitors identified in an ultra-high-throughput screen. *Antimicrobial agents and chemotherapy*, 50(11), 3674-3679.
- [48] Muhammad, M. H., Idris, A. L., Fan, X., Guo, Y., Yu, Y., Jin, X., & Huang, T. (2020). Beyond risk: bacterial biofilms and their regulating approaches. *Frontiers in microbiology*, 11, 928.
- [49] Nadeem, H., Niazi, P., Asif, M., Kaskavalci, G., & Ahmad, F. (2021). Bacterial strains integrated with surfactin molecules of *Bacillus subtilis* MTCC441 enrich nematocidal activity against *Meloidogyne incognita*. *Plant Biology*, 23(6), 1027-1036.
- [50] Nechacov, S. (2019). Factors Affecting Biofilm Formation in Oral Pathogenic Bacteria of the Red Complex.
- [51] Nwaiwu, O., & Aduba, C. C. (2020). An in silico analysis of acquired antimicrobial resistance genes in *Aeromonas* plasmids. *AIMS microbiology*, 6(1), 75.
- [52] O'Toole, G., Kaplan, H. B., & Kolter, R. (2000). Biofilm formation as microbial development. *Annual Reviews in Microbiology*, 54(1), 49-79.
- [53] O'Toole, G. A., Pratt, L. A., Watnick, P. I., Newman, D. K., Weaver, V. B., & Kolter, R. (1999). [6] Genetic approaches to study of biofilms. *Methods in enzymology*, 310, 91-109.
- [54] Patwardhan, S. B., Pandit, C., Pandit, S., Verma, D., Lahiri, D., Nag, M., & Prasad, R. (2022). Illuminating the signalomics of microbial biofilm on plant surfaces. *Biocatalysis and Agricultural Biotechnology*, 102537.
- [55] Pinski, A., Betekhtin, A., Hupert-Kocurek, K., Mur, L. A., & Hasterok, R. (2019). Defining the genetic basis of plant-endophytic bacteria interactions. *International Journal of Molecular Sciences*, 20(8), 1947.
- [56] Prouty, A. M., Schwesinger, W. H., & Gunn, J. S. (2002). Biofilm formation and interaction with the surfaces of gallstones by *Salmonella* spp. *Infection and immunity*, 70(5), 2640-2649.
- [57] Pinski, A., Betekhtin, A., Hupert-Kocurek, K., Mur, L. A., & Hasterok, R. (2019). Defining the genetic basis of plant-endophytic bacteria interactions. *International Journal of Molecular Sciences*, 20(8), 1947.
- [58] Ramey, C. T., & Ramey, S. L. (2004). Early learning and school readiness: Can early intervention make a difference?. *Merrill-Palmer Quarterly*, 50(4), 471-491.
- [59] Rybtke, M., Hultqvist, L. D., Givskov, M., & Tolker-Nielsen, T. (2015). *Pseudomonas aeruginosa* biofilm infections: community structure, antimicrobial tolerance and immune response. *Journal of molecular biology*, 427(23), 3628-3645.
- [60] Shankar, V., Baghdayan, A. S., Huycke, M. M., Lindahl, G., & Gilmore, M. S. (1999). Infection-derived *Enterococcus faecalis* strains are enriched in esp, a gene encoding a novel surface protein. *Infection and immunity*, 67(1), 193-200.
- [61] Sorroche, F. G., Spesia, M. B., Zorreguieta, Á., & Giordano, W. (2012). A positive correlation between bacterial autoaggregation and biofilm formation in native

Sinorhizobium meliloti isolates from Argentina. *Applied and environmental microbiology*, 78(12), 4092-4101.

[62] Tan, M. S., White, A. P., Rahman, S., & Dykes, G. A. (2016). Role of fimbriae, flagella and cellulose on the attachment of Salmonella Typhimurium ATCC 14028 to plant cell wall models. *PLoS One*, 11(6), e0158311.

[63] Trivedi, P., Leach, J. E., Tringe, S. G., Sa, T., & Singh, B. K. (2020). Plant-microbiome interactions: from community assembly to plant health. *Nature reviews microbiology*, 18(11), 607-621.

[64] Trunk, T., Khalil, H. S., & Leo, J. C. (2018). Bacterial autoaggregation. *AIMS microbiology*, 4(1), 140.

[65] Vander Hoogerstraete, T., Wellens, S., Verachtert, K., & Binnemans, K. (2013). Removal of transition metals from rare earths by solvent extraction with an undiluted phosphonium ionic liquid: separations relevant to rare-earth magnet recycling. *Green Chemistry*, 15(4), 919-927.

[66] Van Houdt, R., & Michiels, C. W. (2005). Role of bacterial cell surface structures in Escherichia coli biofilm formation. *Research in microbiology*, 156(5-6), 626-633.

[67] Vasudevan, R. (2014). Biofilms: microbial cities of scientific significance. *J Microbiol Exp*, 1(3), 00014.

[68] Venturi, V., & Bez, C. (2021). A call to arms for cell-cell interactions between bacteria in the plant microbiome. *Trends in Plant Science*, 26(11), 1126-1132.

[69] Waters, C. M., & Bassler, B. L. (2005). Quorum sensing: cell-to-cell communication in bacteria. *Annu. Rev. Cell Dev. Biol.*, 21, 319-346.

[70] Wang, H., Ding, S., Dong, Y., Ye, K., Xu, X., & Zhou, G. (2013). Biofilm formation of Salmonella

serotypes in simulated meat processing environments and its relationship to cell characteristics. *Journal of food protection*, 76(10), 1784-1789.

[71] Wheatley, R. M., & Poole, P. S. (2018). Mechanisms of bacterial attachment to roots. *FEMS microbiology reviews*, 42(4), 448-461.

[72] Wilke, G. A., & Wardenburg, J. B. (2010). Role of a disintegrin and metalloprotease 10 in Staphylococcus aureus  $\alpha$ -hemolysin-mediated cellular injury. *Proceedings of the National Academy of Sciences*, 107(30), 13473-13478.

[73] Wilson, J. W., Schurr, M. J., LeBlanc, C. L., Ramamurthy, R., Buchanan, K. L., & Nickerson, C. A. (2002). Mechanisms of bacterial pathogenicity. *Postgraduate medical journal*, 78(918), 216-224.

[74] Wozniak, S. E., Gee, L. L., Wachtel, M. S., & Frezza, E. E. (2009). Adipose tissue: the new endocrine organ? A review article. *Digestive diseases and sciences*, 54, 1847-1856.

[75] Wu, L., Gao, Y., Liu, J., & Li, H. (2017). Event-triggered sliding mode control of stochastic systems via output feedback. *Automatica*, 82, 79-92.

[76] Wurpel, D. J., Totsika, M., Allsopp, L. P., Hartley-Tassell, L. E., Day, C. J., Peters, K. M., & Schembri, M. A. (2014). F9 fimbriae of uropathogenic Escherichia coli are expressed at low temperature and recognise Gal $\beta$ 1-3GlcNAc-containing glycans. *PloS one*, 9(3), e93177.

[77] Yaron, S., & Römling, U. (2014). Biofilm formation by enteric pathogens and its role in plant colonization and persistence. *Microbial biotechnology*, 7(6), 496-516.